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## PRODUCT DATASHEET



Inert adhesive spacers for the protection of mounted biological structures in light microscopy. For research use only.

Ready-to-use.

**AD-SEAL** are spacers for use in light microscopy. These spacers are designed as a special durable non-transparent layer of inert adhesive with a precisely defined size and thickness.

The spacer precisely defines the space between the cover and the substrate glass or between two covers, depending on the mode of use. This is crucial for protecting the mounted biological structures, as it protects them from both external pressure and capillary pressure between the two glasses, which is most commonly the cause of spreading or flattening of the biological specimen.

With the use of **AD-SEAL** spacers, it is possible to use liquid non-setting mounting media without the need for further fixation with varnish or glue. Proper use of **AD-SEAL** also prevents the release and leakage of mounting media from the space of the microscopic specimen. **AD-SEAL** is produced in black color for quick navigation on the spacer edge during microscopic observation.

**AD-SEAL** spacers are recommended to use together with **AD-MOUNT** non-hardening mounting media. This combination is suitable for mounting bigger biological objects, e.g. oocytes, early embryos or organoids.

**Available package:** 20 pcs (*Cat. No.: ADS-18-10150-20*), 100 pcs (*Cat. No.: ADS-18-10150-100*)

Storage instructions: a temperature range of 4-30°C is recommended

After application resistance: the bond's resistance limits are -40°C to 120°C."

**Dimensions:** 

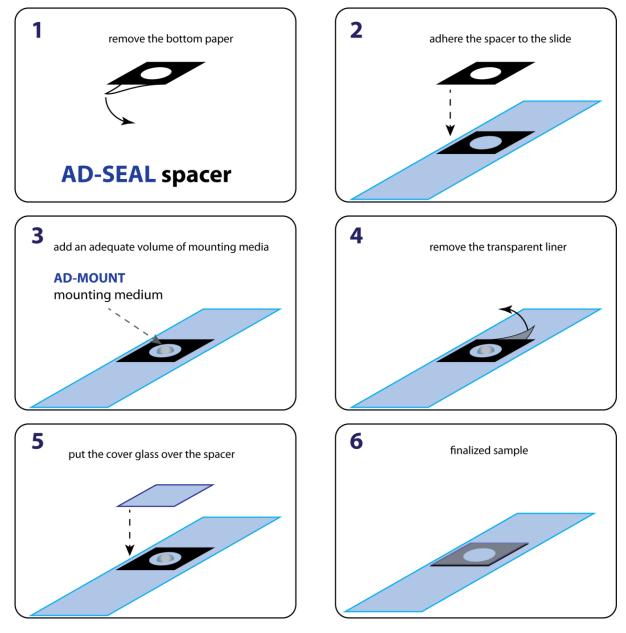
AD-SEAL SQ18-10150

Square side length: 18 mm Inner space diameter: 10 mm Spacer thickness: 0.150 mm



## **Optimized Procedure:**

The illustrated procedure describes the step-wise manner in usage of **AD-SEAL** spacers. The removing the covering paper from the spacer (1), adhering the spacer to the slide (2), adding the mounting media – at this step where the bigger samples like oocytes, organoids are added (3), removing the covering transparent liner (4) and finalizing by placing the cover glass over the spacer (5, 6).



Note: This procedure is a general recommendation and may need to be optimized for individual laboratory procedures.